Morphological Description of a Plankton-Based Megalopa *Thalamita* sp. (Crustacea: Brachyura: Portunidae) from the Obhur Creek, Red Sea, Saudi Arabia

Indra Effendy¹, A. A. J. Kumar¹ and Mohsen M. El-Sherbiny^{1,2}

¹Department of Marine Biology, Faculty of Marine Sciences, King Abdulaziz University, Jeddah, Saudi Arabia, and ² Marine Sciences Department, Faculty of Science, Suez Canal University, Ismailia, Egypt

Abstract. Megalopae of a *Thalamita* sp. crab, collected from the Obhur Creek, Jeddah, Saudi Arabia, are morphologically described. The present specimens are compared with megalopae of two species of this genus: *Thalamita pelsarti* from Japan water and *Thalamita crenata* from Indian waters. Some morphological features are common for megalopae of *Thalamita*: 3-segmented antennal peduncle, 8-segmented antennal endopod, 2-segmented endopod of mandible, 5-segmented endopod of maxilliped III and 3 hooks in the endopod of pleopods I–IV. The morphological differences found between the megalopae of *Thalamita* spp. are found in various appendages of antennule, antenna, maxillule, maxilla, maxillipeds I–III and pleopods.

Keywords: Megalopa, Morphological description, Thalamita, The Red Sea.

1. Introduction

Thalamita species crabs are widely distributed in the Indo-West Pacific regions (Cannicci et al., 1996). Eighty-nine species of Thalamita Latreille, 1829 have been enlisted by Ng et al. (2008). Among those eighty-nine, eleven species are recorded in the Red Sea. The are Thalamita chaptalii Audouin, species 1826, Thalamita mitsiensis Crosnier, 1962, Thalamita crenata Rüppell, 1830, Thalamita danae Stimpson, 1858, Thalamita iranica 1946, Stephensen, Thalamita murinae Zarenkov, 1971, Thalamita poissonii Savigny, 1817, Thalamita prymna Herbst, 1803. Thalamita quadrilobata Miers. 1884. Thalamita savignyi A. Milne-Edwards, 1861, Thalamita sexlobata Miers, 1886 (Apel and 1998; Spiridonov, Spiridonov, 1999; Spiridonov and Neumann, 2008).

Megalopa shares the morphological features of both zoeal and post-larval stages as it is a transitional stage between them (Rice, 1980; Gore, 1985; Cházaro-Olvera, 1996; Cházaro-Olvera and Peterson, 2004). The morphological features of megalopae vary in relation to the differences in the feed and environmental conditions (McConnaughey, 1974; Barnes, 1980; Mc Laughlin, 1980).

Most megalopae described are laboratory-reared and descriptions of oceancollected larvae are very scanty. Descriptions of the latter are important as their morphological features are shaped by natural environmental conditions unlike larvae reared in laboratories, where all such conditions are controlled (Cuesta *et al.*, 2002). Therefore, to have a complete understanding of the larval morphological features, those collected from their natural habitats also ought to be described and such descriptions are very meagre. In the present study, ocean-collected *Thalamita* larvae are described.

Zoeal descriptions of Thalamita sp. are abundant: T. admete Herbst, 1803 (Greenwood and Fielder, 1979), T. crenata Rüppell, 1830 (Chhapgar, 1956; Thomas et al., 1980; Krishnan and Kannupandi, 1990); T. danae Stimpson, 1858 (Fielder and Greenwood, 1979; Jiang et al., 2007); T. pelsarti Montgomery, 1931 (Islam et al., 2005). T. poissoni Auodouin, 1826 (Al-Kholy, 1963), T. prymna Herbst, 1803 (Terada, 1986) and, T. sima H. Milne Edwards, 1934 (Terada, 1979). Morphological descriptions of megalopa are limited to T. crenata Rüppell, 1830 (Thomas et al., 1980; Krishnan and Kannupandi, 1990), T. danae Stempson, 1858 (Fielder and Greenwood, 1979) and Т. pelsarti Montgomery, 1931 (Islam et al., 2005). All these descriptions were made based on laboratory-reared specimens.

In the present study, *Thalamita* sp. megalopae obtained from ocean-collected meroplankton are morphologically described.

2. Material and Methods

Meroplankton were collected at dusk using a plankton net with a mesh size of 150 µm. The net was kept at one meter depth in Obhur Creek for two hours soon after sun set. An Hollis LED 6 underwater torch light with 320 lumens were used to attract the zooplanktons. Altogether, 26 megalopae were obtained. Of those, 18 were portunid megalopae. Six of these were morphologically similar based on pigment distribution on carapace and ambulatories, rostral spine length and width of front and carapace. White dots were thickly distributed on the carapace and ambulatories in those larvae. Four of these larvae were preserved in 70% alcohol, whereas the other two successfully moulted into first crab and were found to belong to the genus *Thalamita.* The exuviae of the megalopae that moulted into first crab were preserved in 70% alcohol. The preserved megalopae and exuviae were dissected in polyvinyl lactophenol with a dissection microscope Leica M80 (Leica Camera AG, Germany). Using the same microscope, measurements of the larval appendages were made. Appendages of the larvae were drawn using a differential interference contrast microscope (Leica DM 6000B) equipped with a camera lucida and setal counts were made. The morphological features of the megalopae and exuviae were compared.

The cephalothorax width (CW, maximum distance across the carapace) and carapace length (CL, distance between the base of the rostral spine and posterior border of the carapace) were measured. Setal counts and morphological descriptions were made following Clark et al. (1998).

3. Results and Discussion

 $CL = 1.2 \pm 0.02$ mm; $CW = 0.83 \pm 0.015$ mm.

Carapace. (Fig. 1A) Longer than wide; smooth, narrowing anteriorly. Rostrum ending in pointed tips projecting anteriorly; dorsal, lateral spines absent; 12–14 simple setae on posterolateral and 8–10 simple setae on posterior margins. Eyes well developed.

Antennule. (Fig. 1B) Peduncle 3segmented, basal segment with simple seta. Endopod with 6 segments (0, 4 aesthetascs, 3 aesthetascs + 2 simple setae, 2 aesthetascs, simple seta, simple seta). Exopod unsegmented with 2 simple setae terminally.

Antenna. (Fig. 1C) Peduncle 3segmented with 1, 2, 1 simple setae. Flagellum 8-segmented with 2, 0, 1, 2, 0, 2, 0, 3 setae.

Mandible. (Fig. 2A) Palp 2-segmented, proximal segment with seta, distal segment with 6 plumose setae, 2 simple setae.

Maxilulle. (Fig. 2B) Protopod with simple long seta. Coxal endite with 3 simple setae, 5 plumose setae, 5 cuspidate setae. Basal endite with 4 simple setae, 4 plumodenticulate, 25 cuspidate setae. Endopod 2-segmented, simple, plumose seta on proximal segment. The second segment with plumose seta, 2 simple terminal setae.

Maxilla. (Fig. 2C) Coxal endite bilobed with 5 + 3 plumose setae, basal endite bilobed with 6 + 6 plumose setae. Scaphognathite with 41 plumose setae.

First maxilliped. (Fig. 3A) Coxal endite with 7 plumose setae, basal endite with 13 simple + 16 plumose setae. Endopod unsegmented with 3 simple setae. Exopod 2segmented with 0, 4 serrated setae. Epipod unsegmented with 4 long simple proximal setae, 3 long simple medial setae, 3 long simple terminal setae.

Second maxilliped. (Fig. 3B) Endopod 4-segmented with 2 simple, 2 simple, 2 simple + 1 plumose + 2 serrated, 5 simple + 3 serrated setae. Exopod 2-segmented with 0, 5 plumose setae.

Third maxilliped. (Fig. 3C) Protopod with 3 simple, 2 plumose, 2 serrated setae. Endopod 5-segmented: 5 simple + 1 plumose + 10 serrated setae, 7 simple + 3 serrated setae, 3 simple + 4 serrated setae, 1 simple + 6 serrated setae, 6 serrated setae.

Pereiopods. (Fig. 4A–C) Cheliped with simple setae on every segment. Dactyli of segments II–IV with plumose setae, propodi of segments II–IV with plumose, simple setae; other segments with simple setae. Distal segment of pereiopod V paddle-shaped with 3 long setae.

Abdomen. (Fig. 5A) Abdomen with 6 somites, each with 2 pairs of simple setae on dorsal surface. Telson broader than long.

Pleopods. (Fig. 5B–F) Pleopods I–IV with 16, 15, 12, 8 plumose setae on exopod, respectively; endopods with three cincinnuli each. Uropod with 9 plumose setae.

The length of the megalopae of *Thalamita* sp. ranged between 1.08 and 1.68 cm and the size was within 1.36 cm.

The first crabs obtained from the two megalopae belonged to *Thalamita* sp. All the morphological features of the dissected larvae were found to be concordant with those of the exuviae of the megalopae and this shows that all the dissected larvae are of the same species of *Thalamita*. The first crabs were not morphologically described as the number of specimens are too low for taxonomic descriptions.

The morphological features of crustacean larvae are shaped by varying prey abundance (Welch and Epifano, 1995). The features of laboratory-reared larvae and those collected from their natural habitats vary considerably as the abiotic and biotic factors, viz. food, salinity and temperature, are not stable in the ocean as in laboratories (Marco-Herrero *et al.*, 2014). Therefore, ocean-collected megalopae are highly needed for taxonomic studies.

Bookhout and Costlow (1974) remarked that the zoeal stages of different species of *Thalamita* are difficult to be distinguished as their morphological features are highly similar. However, the present study revealed that megalopae of different species of *Thalamita* exhibit remarkable morphological variations. The number of segments of the peduncle, exopod and endopod of antennule, peduncle and flagellum of antenna and endopod of maxilliped II varied. Similarly, the setations of the various appendages of antennule, antenna, maxillule, maxilla, maxillipeds I–III and pleopods varied (Table I). Based on these variations, the megalopae of *Thalamita* sp. can be identified from meroplankton.

Morphological variations have also been recorded between megalopae of conspecific *Thalamita* collected from two different regions (Table I): *T. crenata* I collected by Krishnan and Kannupandi (1990) from east coast of India and *T. crenata* II collected by Thomas *et* *al.* (1980) from the west coast of India. Differences appeared in segmentation: endopod of antennule, 2-segmented and unsegmented for *T. crenata* I and II, respectively; exopod of antennule, 4- and 5segmented for *T. crenata* I and II, respectively. Differences were also recorded in setation of *T. crenata* I and II.

 Table 1. Morphological differences between megalopae of 3 species of Thalamita Latreille, 1829. Abbreviations: A, aesthetascs; CL, carapace length; CW, carapace width; Lss, lateral setae; Sch, scaphognathite; S, setae; Unseg, unsegmented.

Characters		Present	<i>T. pelsarti</i> (Islam et al, 2005)	<i>T. crenata</i> (Krishnan and Kannupandi, 1990)	<i>T. crenata</i> (Thomas <i>et al</i> , 1980)
Carapace	CL in mm	1.36	1.08	1.68	1.52
Antennule	CW in mm Peduncle	3 (18, 0, 0)	1.91 3 (3S, 4S, 1S)	1.26 3 (5S, 4S, 2S)	3 (0, 0, 0)
	Endopod Exopod Ae+S	2 (0, 2S) 6 (0, 4A, 3A2S, 2A, 1S, 2S)	Unseg (5S) 5 (0, 4A, 5A, 4A2S, 2S2S)	2 (0, 4S) 4 (1S, 4A, 3A2S, 2A2S)	Unseg (5S) 5 (0, 4A, 4A, 4A, 4S)
Antenna	Peduncle Endopod	3 (0, 1S, 1S) 8 (2S, 0, 1S, 2S, 0, 2S, 0, 3S)	3 (3S, 2S, 3S) 8 (0, 0, 3S, 2S, 4S, 2S, 3S, S)	4 (1S, 2S, 2S, 1S) 8 (0, 0, 3S, 2S, 4S, 2S, 3S, 2S)	4 (0, 0, 0, 0) 8 (0, 0, 0, 0, 2S, 0, 0, 3S)
Mandible	Palp	2 (1S, 8S)	2 (0, 9S)	2 (0, 7S)	2 (0, 7S)
Maxillule	Coxa Basis Endopod	Unseg (13S) Unseg (33S) 2 (2S, 3S)	Unseg (16S) Unseg (24S) 2 (2S, 2S)	Unseg (11S) Unseg (20S) Unseg (3S)	Unseg (23S) Unseg (13S) Unseg (5S)
Maxilla	Protopod Coxa	Unseg (1S) (5+3)S	Unseg (2S) (7 + 7)S	(4+6)S	(5+6)S
	Basis Endopod	(6+6)S	(8 + 10)S Unseg (4)	(7 + 9)S	(6+6)S
N.C. 1111 1	Sch Lss	41S	508	67S	46S
Maxilliped I	Coxa Basis Endopod	7S 29S 3S	14S 27S 4S	13S 30S 7S	12S 27S 4S
	Exopod	2(0, 4S)	2 (1S, 3S)	2 (1S, 5S)	2 (0, 4S)
Maxilliped	Epipod	98	158	128	4S
11	Coxa & Basis				
	Endopod Exopod	4 (2S, 2S, 5S, 9S) 2 (0, 4S)	4 (3S, 1S, 8S, 9S) 2 (1S, 5S)	4 (6S, 2S, 5S, 9S) 2 (1S, 5S)	5 (1S, 2S, 0, 8S, 8S) 2 (0, 5S)
	Epipod				98
Maxilliped III	Coxa & Basis	5 (16S, 10S, 7S, 7S, 6S)	Fused (3S) 5 (23S, 15S, 9S, 9S, 10S)	5 (23S, 15S, 10S, 14S, 11S)	5 (15S, 12S, 6S, 7S, 9S)

	Exopod Epipod	2 (0, 3S) 4S	2 (3S, 6S) 24S	2 (3S, 6S) 17S	2 (5S, 0) 15S
	Protopod Endopodal	58		10S	
Pleopod I	Hook	3	3	3	3
	Exopod Endopodal	168	198	208	158
Pleopod II	Hook	3	3	3	3
	Exopod Endopodal	15S	228	198	155
Pleopod III	Hook	3	3	3	3
-	Exopod Endopodal	12S	20S	19S	15S
Pleopod IV	Hook	3	3	3	3
_	Exopod Endopodal	8S	19S	18S	158
Pleopod V	Hook	Absent	Absent	Absent	Absent
	Exopod	9S	11S	11S	14S
	protopod			1S	1S



Fig. 1. Thalamita sp. megalopa A) Carapace; B) Antennule; C) Antenna.



Fig. 2. *Thalamita* sp. megalopa A) Mandible; B) Maxillule; C) Maxilla.



Fig. 3. Thalamita sp. megalopa A) Maxilliped I; B) Maxilliped II; C) Maxilliped III.



Fig. 4. Thalamita sp. megalopa A) Pereiopod I; B) Pereiopod II; C) Pereiopod III; D) Pereiopod IV; E) Pereiopod V.



Fig. 5. Thalamita sp. megalopa A) Abdomen; B) Pleopod I C) Pleopod II; D) Pleopod III; E) Pleopod IV; F) Uropod.

4. Conclusions

Morphological similarities between Thalamita sp. megalopae are very few: 3segmented antennal peduncle, 8-segmented antennal endopod, 2-segmented endopod of mandible, 5-segmented endopods of maxilliped III, and the number of hooks in the endopod of pleopods I-IV (3). Reiger (1998) pointed out that more morphological data of brachyuran larvae are needed to establish generic and specific characters. Therefore, these similar larval morphological features cannot be considered as generic characters at present.

References

- Al-Kholy, A. A. (1963). Some larvae of decapod Crustacea (from Red Sea). Publ. Mar Biol. Stn., Al-ghardaqa (Red sea), 12: 159-176.
- Apel, M. and Spiridonov, V. A. (1998). Taxonomy and zoogeography of the portunid crabs (Crustacea: Decapoda: Brachyura: Portunidae) of the Arabian Gulf and adjacent waters (pp. 159-331). KSA, Riyadh: National Commission for Wildlife Conservation and Development.
- **Barnes, R. D.** (1980). *Zoología de los invertebrados* (3rd ed.). Interamericana, Mexico.
- Bookhout, C. G. and Costlow, J. D. Jr. (1974). Larval development of *Portunus spinicarpus* reared in laboratory. *Bulletin of Marine Science*, 24: 20-51.
- Cannicci, S., Dahdouh-Guebas, F., Anyona, D. and Vannini, M. (1996). Natural diet and feeding habits of *Thalamita crenata* (Decapoda: Portunidae). *Journal of Crustacean Biology*, 16(4):678-683.
- Cházaro-Olvera, S. (1996). Descripción de las megalopas de las especies Callinectes sapidus (Rathbun), C. similis (Williams), C. rathbunae (Contreras), Arenaeus cribrarius (Lamarck) y Pachygrapsus gracilis (Saussure) de la boca de communicaión de laguna Camaronera, Alvarado. M. Sc. Thesis Facultad de Ciencias, UNAM Mexico Ver.: 1-118.
- Cházaro-Olvera, S. and Peterson, M. S. (2004). Effects of salinity on growth and molting of sympatric *Callinectes* spp. from Camaronera lagoon, Veracruz, Mexico. *Bulletin of Marine Science*, 74(1): 115-127.
- Chhapgar, B. F. (1956). On the breeding habitat and larval stages of some crabs of Bombay. *Records of the Indian Museum*, **54**: 33-52.

- Clark, P. F., Calazans, D. D. and Pohle, G. W. (1998). Accuracy and standardization of brachyuran larval descriptions. *Invertebrate Reproduction and Development*, **33**(2–3): 127–144.
- Cuesta, J. A., Luppi, T. A., Rodríguez, A. and Spivak, E. D. (2002). Morphology of the megalopal stage of *Chasmagnathus granulatus* Dana, 1851 (Crustacea: Decapoda: Brachyura: Varunidae), with comments on morphological anomalies. *Proceedings of the Biological Society of Washington*, 115: 391-402.
- Fielder, D. R. and Greenwood, J. C. (1979). Larval development of the swimming crab *Thalamita danae* Stimpson, 1858 (Decapoda, Portunidae), reared in the laboratory. *Proceedings of the Royal Society of Queensland*, 90: 13–20.
- Gore, H. R. (1985). Molting and growth in decapod larvae. In: A.M. Wenner (ed.), Crustacean Issues 2: Larval Growth. London, Taylor & Francis Groups, pp: 1-65.
- Greenwood, J. G. and Fielder, D. R. (1979). A comparative study of the first and final zoeal stages of four species of *Thalamita* (Crustacea: Portunidae). *Micronesica*, 15: 309-314.
- Islam, S., Machiko, K. and Shigemitsu, S. (2005). Larval development of the swimming crab *Thalamita pelsarti* Montgomery, 1931 (Crustacea: Brachyura: Portunidae) reared in the laboratory. *Russian Journal of Marine Biology*, 31: 78-90.
- Jiang, G. C., Shy, J. Y. and Lai, H. T. (2007). Morphological Observation on the Early Larval Development of *Thalamita danae* (Crustacea, Decapoda, Portunidae). *Journal Fisheries Society of Taiwan*, 34(3): 247-259.
- Krishnan, T. and Kannupandi, T. (1990), Laboratory cultured zoea, megalopa and first crab of the estuarine crab *Thalamita crenata* (Latr.) A.Milne Edwardes 1861 (Brachyura: Portunidae). *Mahasagar*, 23(2): 139-152.
- Marco-Herrero, E., Gonzales-Gordillo, J. I. and Cuesta, J. A. (2014). Morphology of the megalopa of the mud crab, *Rhithropanopeus harrisii* (Gould, 1841) (Decapoda, Brachyura, Panopeidae), identified by DNA barcode. *Helgoland Marine Research*, 68:201-208.
- McConnaughey, H. B. (1974). Introducción a la biología marina. Acribia, Zaragoza pp: 1-459.
- McLaughlin, P. A. (1980). Comparative morphology of recent Crustacea. San Francisco, California. Freeman and Co., pp: 1-177.
- Ng, R. K. L., Guinot, D. and Davie, R. J. R. (2008). Systema Brachyurorum: Part I. An annotated checklist of extant brachyuran crabs of the world. *Raffles Bulletin of Zoology*, 17: 1-286.
- Rice, A. L. (1980). Crab zoea morphology and its bearing on the classification of the Brachyura. *Transaction of the Zoological Society of London*, 35: 271-424.

- Rieger, P. J. (1998). Desenvolvimento larval de Uca (Minuca) burgersi Holthuis (Crustacea, Decapoda, Ocypodidae), em laboratorio. *Revista Brasileira de Zoologia*, 15(3): 727-756.
- Spiridonov, V. A. (1999). Results of the Rumphius Biohistorical Expedition to Ambon (1990). Part 8. Swimming crabs of Ambon (Crustacea: Decapoda: Portunidae). Zoologische Mededelingen, **73**: 63-97.
- Spiridonov, V. A. and Neumann, V. (2008). Coralinhabiting swimming crabs (Crustacea, Decapoda, Portunidae) of the Sudanese Red Sea. Organisms Diversity and Evolution, 8(3): 170-e1.
- Terada, M. (1979). A classification of zoea larvae in the subfamily Portuninae in the family Portunidae. *Zoological Magazine*, 88: 254-268.
- Terada, M. (1986). Zoeal development of the swimming crab Thalamita prymna (Herbst), Portunidae, Portuninae. Researches on Crustacea, 15: 15-23.
- Thomas, M., Khan S. A., Kannupandi T. and Natarajan R. (1980). Laboratory reared larval stages of the Portunid crab *Thalamita crenata* Milne Edwards. *Indian Journal* of Marine Science, 9: 263–270.
- Welch, J. M. and Epifano, C. E. (1995). Effect of variation in prey abundance on growth and development of crab larvae reared in laboratory and in large field-deployed enclosures. *Marine Ecology Progress Series*, 116: 55-64.

الوصف الظاهري ليرقة الميجالوبا لنوع .*Thalamita* sp (القشريات، قصيرات الذنب، السلطعونات السابحة) من خور أبحر، البحر الأحمر، المملكة العربية السعودية إندرا أفندي¹، وأناند جيو كومار¹، ومحسن الشربيني^{2،1}

¹ قسم الأحياء البحرية، كلية علوم البحار ، جدة، المملكة العربية السعودية، و²قسم علوم البحار ، كلية العلوم، جامعة قناة السويس، مصر

Thalamita في هذه الدراسة، تم وصف يرقة الميجالوبا للسلطعون البحري من نوع Thalamita .sp. والتي تم جمعها من خور أبحر، جدة، المملكة العربية السعودية. وتمت مقارنة يرقات *Thalamita والتي تم جمعها من خور أبحر، جدة، المملكة العربية السعودية. وتمت مقارنة يرقات Thalamita الميجالوبا من البحر الأحمر مع نقس اليرقات لنوعين من هذا الجنس، وهما Thalamita الميجالوبا من مياه اليابان و Thalamita crenata من المياه الهندية. وظهرت بعض الصفات المورفولوجية الشائعة في يرقات الميجالوبا لهذا الجنس في عينات البحر الأحمر ، مثل: الساق المورفولوجية الشائعة في يرقات الميجالوبا لهذا الجنس في عينات البحر الأحمر، مثل: الساق المورفولوجية الشائعة في يرقات الميجالوبا لهذا الجنس في عينات البحر الأحمر، مثل: الساق المعقلة لقرون الاستشعار، والشدفة الإنسية (endopod) لقرن الاستشعار مكونة من 8 عقل، والشدفة الإنسية في اللحى (endopod) مكونة من عقلتين، والشدفة الإنسية لأرجل الفكية الثالثة المعقلة القرون الاستشعار، والشدفة الإنسية (endopod) لقرن الاستشعار مكونة من 8 عقل، والشدفة الإنسية في اللحى (endopod) مكونة من عقلتين، والشدفة الإنسية لارجل الفكية الثالثة المعقلة لقرون الاستشعار والشدفة الإنسية (endopod) لقرن الاستشعار مكونة من 8 عقل، والشدفة الإنسية لأرجل العوم (1–4) وتحتوي والشدفة الإنسية لأرجل العوم (1–4) وتحتوي على بعض الاختلافات المورفولوجية بين يرقة الميجالوبا لنوع علي ثلاثة خطافات. تم العثور على بعض الاختلافات المورفولوجية بين يرقة الميجالوبا لنوع المي ثلاثة خطافات. والأخرى في عدد الزوائد المختلفة، مثل قرن الاستشعار الأول والثاني، علي ثلاثة خطافات. والفك (maxilliped I-1)، والأرجل الفكية الأولى حتي الثالثة (maxilliped I-1)، وأرجل العوم.*

الكلمات المفتاحية: يرقة الميجالوبا، الوصف الشكلي، Thalamita، البحر الأحمر .